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Endocrine and reproductive parameters in sick and healthy primiparous and multiparous dairy cows



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ABSTRACT

To investigate the association of health status and parity with hormone profiles during the transition period and reproductive parameters in Holstein dairy cows, a prospective observational cohort study was carried out including only healthy primiparous (PP, n=116) and multiparous (MP, n=172) cows at the beginning of the study. A subset of 120 healthy and sick cows was randomly selected for insulin, IGF-I, leptin and adiponectin determination. Primiparous cows had greater IGF-I and adiponectin concentrations (P < 0.05) and tended (P=0.07) to have greater insulin concentrations than MP cows. While healthy and sick MP and sick PP cows presented a sharp decrease in IGF-I concentrations after calving, healthy PP cows maintained them. Postpartum adiponectin concentrations were lower in sick than in healthy MP cows. A greater percentage of healthy cows ovulated during the first 7 weeks after calving when compared to sick cows (67.9% vs 50%, P=0.002) and a similar trend was found for MP vs PP cows (64% vs 53%, P=0.01). More healthy cows were inseminated in comparison to sick cows (94% vs 76.5%, P < 0.01) and more PP than MP cows (90.4% vs 82.7%, P < 0.05). Similarly, healthy cows presented a greater proportion of pregnancy than sick cows (75% vs 54%, P < 0.01) and the proportion of pregnancy was higher in PP than in MP cows (74% vs 56%, P=0.04). Health status interacting with parity yielded different endocrine profiles, which may partially explain the differences in reproductive performance.

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1. Introduction

Due to intensive genetic selection in addition to improvements in nutrition and management of dairy herds, milk production has increased substantially over the last decade [1]. This increase in milk production demands a correct nutritional balance in order to fulfill lactation requirements despite the decrease in voluntary dry matter intake (DMI) around calving [2,3]. The uncoupling of the somatotrophic axis (high growth hormone [GH] vs low insulin-like growth factor-I (IGF-I) concentrations), the increase in hepatic gluconeogenesis, lower glucose utilization by peripheral tissues,

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and lipolysis are homeorhetic adaptations for increasing energy substrates availability [4,5].

Also, the interrelationship of the endocrine, metabolic and immune system may induce a depressed immune system around calving by activating local and systemic defense mechanisms that induce inflammation [6]. This increases the chances of getting sick as it has been shown that 30%–50% of the cows are affected by some form of metabolic or infectious disease around calving [7–9] in addition to reproductive problems or/and increased culling rates [6,10,11]. Additionally, it is known that peripartum diseases are interrelated; e.g., retained placenta to hypocalcemia [12] (which is considered a gateway disease [13]) predisposing an animal to metritis and mastitis [14,15] or affecting reproductive performance [16,17]. Indeed, reproductive failure is still an important reason for involuntary culling in dairy herds [18].

Furthermore, parity affects metabolic adaptation to lactation; primiparous cows, which have not reached their adult body size

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and continue to grow during pregnancy and lactation, present metabolic differences compared to older cows [19]. The competing demands of the mammary gland are superimposed on growth requirements, and both insulin and IGF-I stimulate growth. The profiles of these anabolic hormones and metabolites (such as nonesterified fatty acids [NEFA]) during the transition period according to parity have been inconsistent [20,21]. IGF-I and insulin concentrations are low around calving (because of homeorhetic adaptaaffecting the reproductive axis at both central (hypothalamus-pituitary gland) and peripheral (gonads, reproductive tract, and embryo) level [22]. Indeed, a positive relation between IGF-I and the first postpartum ovulation has been demonstrated [23,24]. Moreover, low IGF-I concentrations were reported in cows developing postpartum diseases, but because of the limited number of cows used, the authors concluded that a reliable association could not be established [25].

Adiponectin and leptin, because of their insulin sensitizing actions and association with body condition, could be potential regulators of metabolism during the transition from pregnancy to lactation [26]. Even if no relationship between leptin and the first postpartum luteal activity could be demonstrated, higher leptin concentrations were associated with shorter intervals to the first observed estrus [27]. Adiponectin is not only a metabolic hormone, as has also been linked to inflammation. Indeed, low adiponectin concentrations around calving have been proposed to be linked to inflammatory disease pathophysiology [28] and could be another mechanism that contributes to poor fertility in dairy cows [26]. As far as we know, there are no reports on endocrine (insulin, IGF-I, leptin and adiponectin) profiles during the transition period in sick and healthy primiparous and multiparous cows and their association with reproductive success.

Taking these findings into account, the objectives of this study were to assess whether health status interacted with parity affecting hormone profiles (IGF-I, insulin, adiponectin, and leptin) during the transition period, and if they were associated with reproductive performance in primiparous (PP) and multiparous (MP) Holstein cows.

2. Materials and methods

2.1. Cows and herd management

The study was conducted in a commercial dairy free stall (loose-housing system) herd in Rio Grande do Sul, southern Brazil. All procedures were carried out in accordance with regulations of the Animal Experimentation Committee (SIPPEE 10.059.15 CEUA 0522017 UNIPAMPA, Brazil).

From a 700-cow herd, 288 Holstein dairy cows ($n=116\,PP$ and $n=172\,MP$) were selected, with an approximate milk yield of 8000 kg per lactation and two calving seasons; spring/summer (October 2014 to January 2015) and autumn/winter (May to August 2015).

Cows were kept on paddocks without significant pasture allowance from day -21 until calving, separated by parity, and were offered 13.3 kg/d of DM as total mixed ration (TMR) including anionic salts according to close-up requirements (NRC, 2001) twice a day at 8:00 and 16:30 hs. After calving, all cows were kept together in a compost barn until 3 days in milk (DIM) and were fed a TMR 3 times daily for *ad libitum* intake in collective feeders, at 5:30, 10:00 and 15:00 hs, and were milked 2 times daily. They did not have access to pasture. The total daily offer of DM was 21.5 kg/d in the spring calving season (2014) and 28.9 kg/d in the autumn calving season (2015). Close-up and fresh cow diet composition has been reported previously [9]. Cows had *ad libitum* access to water and they received 300 mL of propylene glycol, 250 g of calcium

propionate diluted in 1L water orally and 40 g Calcium gluconate subcutaneously at calving. After 3 DIM, cows were translated to a *free-stall* and were fed the same TMR, but milked 3 times daily (DeLaval rotary 80 stations). Data for 305-d milk yield were obtained from DairyPlan C-21 software (GEA).

2.2. Study design, disease recording and sample collection

This is a prospective observational cohort study and only healthy cows were enrolled at the beginning of the study. Peripartum diseases were diagnosed by a single trained veterinarian as reported before [9], using the following criteria: Clinical hypocalcemia was defined as any recumbent cow within 72 hs after parturition exhibiting anorexia, nervous symptoms, staggering, varying degrees of unconsciousness, and good response to intravenously administered calcium. Clinical mastitis was characterized by the presence of abnormal milk or by signs of inflammation in 1 or more quarters, evaluated by the milking person from calving to 30 DIM at the start of milking. Retained placenta (RP) was defined as failure to expel the placenta within 24 h after parturition. Between 3 and 21 \pm 1 DIM, all cows were monitored for metritis twice weekly, every Tuesday and Friday by a veterinarian, using a manual vaginal examination. Presence of abnormal vaginal secretion associated with blood haptoglobin (Hp) concentration > 1 mg/dL, was used for diagnosis of metritis. Lameness was diagnosed weekly using the scale from 1 to 5, where grade 1 had no alteration in gait and at grade 5, the cow was severely limping, without supporting the member on the floor. Cows with lameness were considered those with locomotion score >3. Left displaced abomasum (LDA) was diagnosed based on an auscultation of a "ping" during percussion of the left side of the abdomen. During these health checks, the presence of any other clinical disease(s) was recorded, as well as deaths and culling because of health problems.

Cows were classified according to their health status in healthy cows (without clinical events) and sick cows (one or more clinical events) in order to analyze endocrine and reproductive variables. From the 288 cows evaluated, 45.8% (n = 132) became sick and 1.9% (n = 6) were culled or died within the first 30 DIM. Health status was not affected by parity and the proportions of healthy and sick cows were for MP: 52.9% and 47.0% respectively and for PP: 56.0% and 43.9% respectively.

2.3. Reproductive management

From 50 days after calving, cows were managed with a timed artificial insemination (AI) synchronization protocol consisting in 2 mL of estradiol benzoate (Sincrodiol, Ourofino Saúde Animal Ltda, Brazil), 1 mL of gonadotropin-releasing hormone (GnRH Gestran, ARSA S.R.L, Buenos Aires, Argentina) and an intravaginal progesterone (P4) device (CIDR, Zoetis, Brazil), followed seven days later by an injection of 2 mL prostaglandin (Estron Agener Uniao, Brazil), removing at this time the intravaginal device, and an injection of 0.5 mg estradiol cipionate (ECP, Zoetis, Brazil) [29]. All cows were inseminated 48 h later (47–50 h). Pregnancy diagnosis was performed on day 45 by ultrasound (DVU60vet, Oxson Technology, Brazil). Non-pregnant cows were submitted to the same synchronization protocol.

2.4. Blood sample collection and analyses

From week -2 until week +7, blood samples were collected from the coccygeal vein into 10-mL sterile heparinized tubes. Samples were centrifuged at 3000 x g for 20 min and plasma was stored frozen (-20 °C) until further analysis. In all cows (n = 288) plasma P4 concentrations were measured weekly after calving

using a solid-phase ¹²⁵I radioimmunoassay (RIA) kit (MP, Biomedicals, ICN Biomedicals, Inc. California). The sensitivity of the assay was 0.01 ng/mL, and the intra and inter-assay coefficients of variation (CVs) were not greater than 10.6%. Days to first ovulation (resumption of ovarian cyclicity) was defined as the interval between calving and the first day in which plasma P4 concentrations were >1 ng/mL.

To analyze the endocrine profiles and their relationship with reproductive parameters, a subset of 120 cows that included 57 PP cows (healthy n = 28, sick n = 29) and 63 MP cows (healthy n = 28, sick n = 35) was randomly selected. Insulin, IGF-I, leptin, and adiponectin concentrations were determined on weeks -2, -1, +1, +3, and +5 related to calving (a total of 600 samples for each hormone). IGF-I and insulin concentrations were measured using immunoradiometric assays (IRMA) with commercial kits. Plasma IGF-I concentrations were determined using the IGF-I RIACT kit (Cis Bio International, GIF SUR YVETTE CEDEX, France). The assay's sensitivity was 16 ng/mL, and the intra- and inter-assay CVs for control 1 (47.5 ng/mL) were 8.2% and 8.5%, respectively, and for control 2 (429 ng/mL) 9.9% and 11.9%, respectively. Plasma insulin concentrations were determined using the INS-IRMA kit (DIA Source Immune Assays S.A., Belgium). The assay's sensitivity was 1.3 µIU/mL, and the intra- and inter-assay CVs for control 1 (19.4 µIU/mL) were 5.5% and 8.6%, respectively, and for control 2 (65.6 μ IU/mL) 4.4% and 4.9%, respectively. Leptin concentrations were determined with a liquid phase RIA using a commercial Multi-Species Leptin kit (RIA kit, Millipore, USA). The RIA had a sensitivity of 2.8 ng/mL, and the intra- and inter-assay CVs for control 1 (8.3 ng/mL) were 9% and 10.2% respectively and for control 2 (32 ng/mL) 8.7% and 11% respectively. Adiponectin concentrations were measured using an RIA kit (HADP-61 HK, Millipore, USA) with undiluted plasma samples. The assay's sensitivity was 1.54 ng/mL, and the intra and inter-assay CVs for control 1 (8.4 ng/mL) were 9.1% and 8.5% respectively and for control 2 (81.5 ng/mL) 7.9% and 11.1% respectively.

2.5. Statistical analysis

Insulin, IGF-I, leptin, and adiponectin concentrations were analyzed as repeated measures. The model included the fixed effects of parity (PP vs MP), health status (healthy and sick cows), week, and these factors' interactions. The associations between parity, health status, calving seasons and resumption of ovarian cyclicity, AI, and pregnancy rates were analyzed using 2×2 contingency tables generated by the PROC FREQ statement in SAS (9.2; SAS Institute, 2009). From these tables, the Mantel- Haenszel chisquare test was used to determine the relationship's type 1 error risk. Reproductive parameters were also evaluated by Proc Genmod, including parity and health status in the model. Also, Kaplan-Meier survival analysis for probability of resumption of ovarian cyclicity and pregnancy were performed. Data were further evaluated using multivariable logistic regression (MLR) analysis for resumption of ovarian cyclicity and pregnancy rates. Variables that were not significant (P > 0.05) were removed by manual backward stepwise elimination. Odds ratio (OR) and their respective 95% confidence interval (CI) determined by MLR were used to describe the level of association between the endocrine concentrations, health status, parity and reproductive parameters. Data are reported as least squares means and pooled standard errors. For all statistical analysis P < 0.05 was considered a significant effect, and $P \leq 0.1$ as a trend.

The present study was performed to investigate the role of health status and parity on hormone profiles during the transition period and their association with reproductive performance. The sample size calculation for hormone profiles, assuming a SD of 30%, power = 80%, P-value = 0.05, revealed that 24 animals per group are needed to detect a 25% difference in the mean between two compared groups. For percentage of resumption of ovarian cyclicity in the first 7 weeks after calving, assuming 50 vs 70%, with a power = 80% and P-value = 0.05, 93 animals per group are needed (considering a power = 65%, 66 animals are needed). For pregnancy proportion, assuming 65 vs 80%, with a power = 80% and Pvalue = 0.05, 138 animals per group are needed (the same with a power = 65%, 98 animals are needed). Thus, the sample size considered in this study for endocrine profiles was appropriate, while for reproductive parameters, only moderate differences could be detected.

3. Results

The significance of the fixed effects of the subset groups' hormonal concentrations (n=120) is shown in Table 1. IGF-I and adiponectin concentrations were significantly affected by parity; insulin tended to be affected by this factor, while leptin did not reach significance. The evolution of IGF-I and insulin concentrations according to parity and week differed, and a triple interaction parity*status*week was found for IGF-I.

Insulin concentrations tended to be greater in PP than MP $(19.5 \pm 0.7 \text{ vs } 17.9 \pm 0.6 \text{ uUI/mL}, P = 0.07)$. In PP cows, insulin concentrations remained constant from week -2 until week +5, while in MP cows, insulin concentrations decreased from the closeup period until week +1 (P < 0.003), increasing thereafter until the end of the study without differences according to health status (Fig. 1 A, B). Overall, IGF-I concentrations were higher in PP than MP cows (136.2 \pm 5.3 vs 103.6 \pm 5.0 ng/mL, P < 0.0001). Regardless of health status, IGF-I concentrations increased in PP cows from week -2 to week -1 (P < 0.01) and decreased thereafter until week+1 (P < 0.01) with a sharp decrease in sick PP cows (Fig. 1 C). Although concentrations decreased in all MP cows at week+1, healthy MP cows tended to have greater IGF-I concentrations than sick MP cows (P = 0.1). After week+1, IGF-I concentrations in MP cows tended to increase until week +5, but the close-up concentration was not recovered (Fig. 1 D).

Adiponectin concentrations were greater in PP than MP cows $(21.9\pm1.1\ \text{versus}\ 19.0\pm1.0\ \text{ng/mL},\ P=0.04)$. In all cows, the concentration decreased from week -2 until the end of the study regardless of parity. Also, healthy MP cows tended to have greater adiponectin concentrations than sick MP cows throughout the fresh cow period (P < 0.10, Fig. 1F). Parity did not reach significance for leptin concentrations $(16.4\pm1.4\ \text{and}\ 13.6\pm1.4\ \text{ng/mL}$ for MP

Table 1F-tests of fixed effects for hormone concentrations in cows from week -2 until week +5 according to calving in primiparous (n = 57) and multiparous (n = 63) Holstein cows. Fixed effects were parity, health status (Health), week according to calving and their interactions.

	Parity	Week	Health	Health*week	Parity*week	Health*parity*week
Insulin	0.07	<.0001	0.49	0.94	<.0001	0.74
IGF-I	<.0001	<.0001	0.22	0.01	<.0001	0.03
Adiponectin	0.04	<.0001	0.17	0.19	0.18	0.64
Leptin	0.15	<.0001	0.65	0.89	0.71	0.94

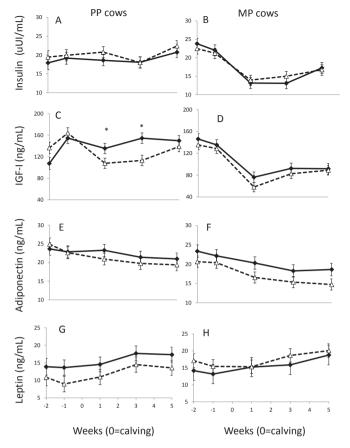


Fig. 1. Insulin (A, B), IGF-I (C, D), adiponectin (E, F) and leptin (G, H) concentrations in Holstein primiparous (PP, n = 57) and multiparous (MP, n = 63) cows from -2 to +5 weeks related to calving according to health status [healthy cows (\bullet) and sick cows (Δ)].

and PP cows, respectively, P = 0.15), and concentrations increased after calving (P < 0.05).

During the first seven weeks after calving, from a total number of 288 cows, 59.7% became cyclic, while 40.3% did not. Health status and parity affected the resumption of ovarian cyclicity (P < 0.05) independent of the calving season. A greater percentage of healthy cows reinitiated ovarian cyclicity when compared to sick cows (67.9% [106/156] vs 50% [66/132], P = 0.002). Although a greater percentage of MP cows reinitiated ovarian cyclicity when compared to PP cows (64% [110/172] vs 53% [62/116], P = 0.01), this difference was due to sick cows (Table 2). Indeed, no differences according to parity were found in healthy cows, but sick PP cows presented a lower probability of resumption of ovarian cyclicity than sick MP cows (Fig. 2 A, B).

In both categories, a greater percentage of healthy cows were inseminated when compared to sick cows (94.0% vs 76.5%, P < 0.01) and this was also greater for PP than MP cows (90.4 vs 82.7,

P < 0.05) (Table 2). The probability of pregnancy was also affected by health status and parity (P < 0.004, Fig. 2 C, D). More healthy cows became pregnant when compared to sick cows (75% vs 54%, P < 0.01) and more PP than MP cows (74% vs 56%, P = 0.04). Conception rate was higher in PP than MP cows (P = 0.01) and in healthy than in sick cows (P = 0.04). Interestingly, while no differences according to health status were found in conception rates of PP cows, sick MP cows had a lower fertilization and/or higher embryonic mortality than healthy MP cows (Table 2).

To analyze the endocrine variables that could explain the reproductive parameters, a logistic regression (LR) was performed including parity, health status, and hormone concentrations; only parity and health status remained in the model (P < 0.05) for AI and pregnancy variables. For resumption of ovarian cyclicity, parity, health status, insulin and IGF-I concentrations remained in the final model (P = 0.01). Insulin concentrations at week -1 were significant for resumption of ovarian cyclicity (OR [IC]); 1.09 [1.01-1.17]) and IGF-I concentration at week +1 (1.01 [1.00-1.02]) (P < 0.05).

4. Discussion

This is the first study associating health status and parity with insulin, IGF-I, leptin and adiponectin profiles during the transition period, and investigating their relation with reproductive parameters. A greater proportion of healthy cows reinitiated ovarian cyclicity during the first 7 weeks after calving compared to sick cows, and the findings for the proportion of insemination and pregnancy were similar. While a lower proportion of PP cows reinitiated their ovarian cyclicity, they presented a greater proportion of pregnancy. Moreover, the significant interaction of health status and parity with IGF-I profiles is novel, and may explain the different reproductive outcome found in healthy and sick primiparous and multiparous cows.

Due to their impact on the reproductive status, endocrine profiles during the transition period were determined [6,11,23]. As a consequence of the increased requirements for milk production and decreased DMI, insulin levels decreased [23,30]. The higher overall IGF-I and insulin concentrations found in PP dairy cows during the transition period have been reviewed previously [19]; milk production is subordinate to growth in PP cows, and these anabolic hormones reveal the difference in metabolic adaptation according to parity. Indeed, the sharp decrease in IGF-I concentrations found in MP cows after calving in contrast to PP cows is an argument for a greater uncoupling of the somatotrophic axis in favour of milk production as previously reported [19]. Adiponectin concentration was also affected by parity and was greater in PP cows. No reports of adiponectin concentrations according to parity in dairy cows have been found, but data are consistent with insulin and IGF-I profiles. Indeed, greater adiponectin concentrations have been related to insulin sensitivity [26]. The adiponectin decrease found after calving is consistent with its role in gluconeogenesis suppression [31], which may facilitate an increase in glucose supply to the mammary gland [32]. Leptin was not affected by parity and

Table 2Reproductive parameters in healthy and sick lactating primiparous (PP) and multiparous (MP) dairy cows.

Variable	PP (n = 116)		MP (n = 172)	
	Healthy (n = 65)	Sick (n = 51)	Healthy (n = 91)	Sick (n = 81)
Resumption of ovarian cyclicity (%)	66 ^a	35 ^b	69 ^a	59 ^c
AI (%)	97 ^a	82 ^b	92 ^a	71 ^b
Pregnancy (%)	82 ^a	66 ^b	69 ^b	43 ^c
Conception (%)	85 ^a	80 ^{ab}	74 ^b	60 ^c

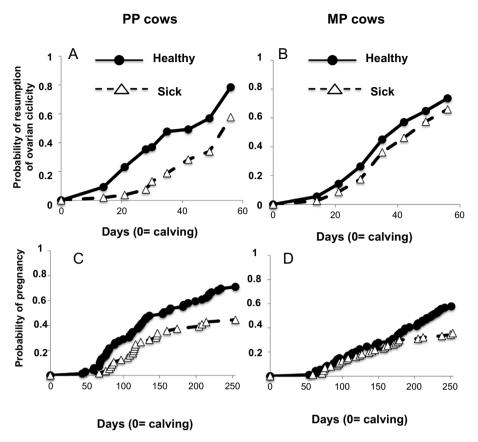


Fig. 2. Probability of resumption of ovarian cyclicity and pregnancy in Holstein primiparous (PP, n = 116) and multiparous (MP, n = 172) cows according to health status [healthy cows (\bullet) and sick cows (Δ)].

concentrations increased after calving in all cows independent of parity, which could be explained by the high concentrate content of the diet in the present study as reported previously [33].

Healthy PP cows maintained greater IGF-I concentrations during the first month after calving, while sick PP cows presented a sharp decrease in response to illness, and this effect was not observed in MP cows. The sharper IGF-I decrease around calving in sick cows was probably associated with a decrease in DMI as previously reported [34,35]. Data suggest a different metabolic adaptation to illness and lactation according to parity. Interestingly, while sick MP cows had greater NEFA concentrations than healthy MP cows, this was not found in PP cows [9]. Thus, the reason for the lower IGF-I in sick PP cows could be the result of calving-induced stress in addition to management and milking routine adaptation at this time [36], and/or a milder NEB associated with a lower milk production when compared to sick MP cows.

Although there was no effect of health status on adiponectin and leptin profiles during the transition period, sick MP cows tended to have lower adiponectin concentrations than healthy cows after calving. Indeed, hypoadiponectinemia has been linked to inflammatory disease pathophysiology [37]. Also, the lower adiponectin concentrations found in sick MP cows after calving are consistent with the lower IGF-I concentrations in this group. Moreover, adiponectin, because of its insulin sensitizing actions, could be a metabolic regulating factor during the transition period [38].

Although the sample size of this study is limited, the reproductive parameters (resumption of ovarian cyclicity, pregnancy and conception rates) were affected by parity. Until day 50 after calving, PP cows presented a lower percentage of resumption of ovarian cyclicity in accordance with previous reports [39]. In dairy cows,

the first ovulation after calving has been linked with the severity and duration of NEB [23]. In several studies [19,21,39,40] the NEB in PP cows has been reported to be at its worst one week before calving and early thereafter. However, the lower NEFA and greater IGF-I concentrations in PP cows (this study and [9]) reflect a better energy balance in this category, so that parity-associated factors mentioned before and reported previously [36] could explain the delay of resumption of ovarian cyclicity in PP cows. In contrast, the proportion of pregnant cows was greater in PP than MP cows. Similarly, Santos et al. [39] reported that MP cows were more likely to start ovarian cyclicity earlier than PP cows, but pregnancy rate was higher in PP cows. In our study, the greater milk production of MP cows could explain the lower proportion of pregnancy because of the increased catabolic state, as milk yield was negatively associated with the ability of lactating cows to conceive and maintain pregnancy [39]. However, others reported no association between milk yield and conception or embryonic survival [39,41]. An increase in liver flux is needed to support higher milk production in MP cows, leading to a greater P4 clearance; thus, lower circulating P4 concentrations could explain embryonic loss and the lower pregnancy proportion in this category [42,43].

Health status affected the percentage of resumption of ovarian cyclicity during the first 7 weeks after calving and was lower in sick cows, which is associated with the worse NEB (lower cholesterol, albumin and IGF-I concentrations) of them ([9] and the present study). Sick cows presented lower DMI and lower cholesterol and albumin concentrations [44,45]. As mentioned before, IGF-I stimulates bovine follicular cells and ovulation [46]. Indeed, IGF-I concentrations one week after calving were associated with resumption of ovarian cyclicity. Beam and Butler [47] also reported

greater IGF-I concentrations two weeks after calving in cows that reinitiated ovarian cyclicity in comparison to non-ovulating cows. Interestingly, the negative effect of illness on resumption of ovarian cyclicity in PP cows was more marked than in MP cows, consistent with the sharper decrease in IGF-I profiles observed in sick PP cows. The lower probability of resumption of ovarian cyclicity in sick cows was consistent with the lower proportion of sick cows that became pregnant. This could be explained by the carryover effects of inflammatory diseases after calving, because disease can induce fever and compromise reproduction, by disrupting oocyte and embryonic development and uterine function which can last for months after the acute diseases has ceased [48]. Interestingly, the interaction of parity and health status with conception rates was significant. Despite a slight numerically difference, in favour of healthy cows, no differences according to health status were found in conception rates in PP cows, although it should be taken into account that for these small differences the sample size is underpowered. Nevertheless, in MP, differences were detected, sick cows had lower conception rates than healthy cows; this may have been due to lower fertilization and/or higher embryonic mortality in sick MP cows. Overall, data suggest that when PP cows overcome the clinical event and start to cycle, their fertility is higher than MP cows. In contrast, in MP cows, greater negative and/or carryover effects of diseases on reproduction were observed, suggesting that fertilization and maintenance of pregnancy are important barriers for reproductive success in this category. Nevertheless, the interpretation of the present data should be carefully performed due to the limited number of cows included for reproductive outcomes.

5. Conclusion

Healthy and sick cows had different endocrine profiles during the transition period, which may partially explain the observed reproductive performance. Parity interacted with health status on reproductive performance; sick PP cows showed a delay in the resumption of ovarian cyclicity and lower IGF-I concentrations, while sick MP cows showed a lower proportion of pregnancy.

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References

- [1] Inguartsen KL. Feeding and management related diseases in the transition cow. Physiological adaptations around calving and strategies to reduce feeding-related diseases. Anim Feed Sci Technol 2006;126:175–213.
- [2] Grummer RR. Impact of changes in organic nutrient metabolism on feeding the transition dairy cow. J Anim Sci 1995;73:2820–33.
- [3] Ingvartsen KL, Andersen JB. Integration of metabolism and intake regulation: a review focusing on periparturient animals. J Dairy Sci 2000;83:1573–97.
- [4] Bell AW. Regulation of organic nutrient metabolism during transition from late pregnancy to early lactation. J Anim Sci 1995;73:2804–19.
- [5] Drackley JK. Biology of dairy cows during the transition period: the final frontier? J Dairy Sci 1999;82:2259–73.
- [6] Esposito C, Irons PC, Webb EC, Chapwanya A. Interactions between negative energy balance, metabolic diseases, uterine health and immune response in transition dairy cows. Anim Reprod Sci 2014;144:60–71.
- [7] LeBlanc S. Monitoring metabolic health of dairy cattle in the transition period. J Reprod Dev 2010;56:29–35.
- [8] Trevisi E, Amadori M, Archetti I, Lacetera N, Bertoni G. Inflammatory response and acute phase proteins in the transition period of high-yielding dairy cows. In: Veas F, editor. Acute phase proteins as early non-specific biomarkers of human and veterinary diseases, Croatia; 2011. p. 355–79.
- [9] Ruprechter G, Adrien ML, Larriestra A, Meotti O, Batista Ch, Meikle A, Noro M. Metabolic predictors of peri-partum diseases and their association with parity in dairy cows. Res Vet Sci 2018;118:191–8.
- [10] Butler ST, Marr AL, Pelton SH, Radcliff RP, Lucy MC, Butler WR. Insulin restores

- GH responsiveness during lactation-induced negative energy balance in dairy cattle: effects on expression of IGF-I and GH receptor 1A. J Endocrinol 2003;176:205–17.
- [11] Bertoni G, Trevisi E, Lombardelli R. Some new aspects of nutrition, health conditions and fertility of intensively reared dairy cows. Ital J Anim Sci 2009;8:491–518.
- [12] Mulligan FJ, Doherty ML. Production diseases of the transition cow. Vet J 2008;176:3—9.
- [13] Goff JP. The monitoring, prevention, and treatment of milk fever and subclinical hypocalcemia in dairy cows. Vet J 2008;176:50–7.
- [14] Curtis CR, Erb HN, Sniffen CJ, Smith RD, Powers PA, Smith MC, White ME, Hillman RB, Pearson EJ. Association of parturient hypocalcemia with eight periparturient disorders in Holstein cows. J Am Vet Med Assoc 1983;183: 559–61.
- [15] Martinez N, Risco CA, Lima FS, Bisinotto RS, Greco LF, Ribeiro ES, Maunsell F, Galvão K, Santos JEP. Evaluation of peripartal calcium status, energetic profile, and neutrophil function in dairy cows at low or high risk of developing uterine disease. J Dairy Sci 2012;95:7158–72.
- [16] Chapinal N, Carson M, LeBlanc SJ, Leslie KE, Godden S, Capel M, Santos JE, Overton M, Duffield TF. The association of serum metabolites in the transition period with milk production and early-lactation reproductive performance. | Dairy Sci 2012;95:1301–9.
- [17] Ribeiro ES, Lima FS, Greco LF, Bisinotto RS, Monteiro AP, Favoreto M. Prevalence of periparturient diseases and effects on fertility of seasonally calving grazing dairy cows supplemented with concentrates. J Dairy Sci 2013;96: 5682–97.
- [18] Gröhn Y, Rajala-Schultz J, Allore HG, DeLorenzo MA, Hertl JA, Galligan DT. Optimizing replacement of dairy cows: modeling the effects of diseases. Prev Vet Med 2003:61:27–43.
- [19] Wathes DC, Cheng Z, Bourne N, Taylor VJ, Coffey MP, Brotherstone S. Differences between primiparous and multiparous dairy cows in the interrelationships between metabolic traits, milk yield and body condition score in the periparturient period. Domest Anim Endocrinol 2007;33:203–25.
- [20] Vandehaar MJ, Yousif G, Sharma BK, Herdt TH, Emery RS, Allen MS, Liesman JS. Effect of energy and protein density of prepartum diets on fat and protein metabolism of dairy cattle in the periparturient period. J Dairy Sci 1999;82: 1282–95.
- [21] Meikle A, Kulcsar M, Chilliard Y, Febel H, Delavaud C, Cavestany D, Chilibroste P. Effects of parity and body condition score at calving on endocrine and reproductive parameters of the dairy cow under grazing conditions. Reproduction 2004;127:727–37.
- [22] Meikle A, de Brun V, Carriquiry M, Soca P, Sosa C, Adrien ML, Chilibroste P, Abecia JA. Influences of nutrition and metabolism on reproduction of the female ruminant. Anim Reprod 2018;15(1):899–911.
- [23] ButlerButler WR. Nutritional interactions with reproductive performance in dairy cattle. Anim Reprod Sci 2000;60–61:449–57.
- [24] Kadivar A, Ahmadi MR, Gheisari HR, Nazifi S. Assessment of IGF-I as a factor influencing postpartum reproductive performance and ovarian condition in dairy cattle. Comp Clin Pathol 2012;21:589–96.
- [25] Piechotta M, Sander AK, Kastelic JP, Wilde R, Heppelmann M, Rudolphi B, Schuberth HJ, Bollwein H, Kaske M. Short communication: prepartum plasma insulin-like growth factor-I concentrations based on day of insemination are lower in cows developing postpartum diseases. J Dairy Sci 2012;95:1367–70.
- [26] Kalamaras K. The role of adiponectin in regulation of metabolism in dairy cows. PhD thesis. University of Nottingham; 2012. Access from the University of Nottingham repository: http://eprints.nottingham.ac.uk/12588/1/ ADIPONECTIN_IN_DAIRY_COWS_KALAMARAS_2012.pdf.
- [27] Liefers SC, Veerkamp RF, Te Pas MF, Delavaud C, Chilliard Y, van der Lende T. Leptin concentrations in relation to energy balance, milk yield, intake, live weight, and estrus in dairy cows. J Dairy Sci 2003;86:799–807.
- [28] Kabara E, Sordillo LM, Holcombe S, Contreras GA. Adiponectin links adipose tissue function and monocyte inflammatory responses during bovine metabolic stress. Comp Immunol Microbiol Infect Dis 2014;37:49–58.
- [29] Pereira MHC, Wiltbank MC, Barbosa LFSP, Costa Jr WM, Carvalho MAP, Vasconcelos JLM. Effect of adding a gonadotropin-releasing-hormone treatment at the beginning and a second prostaglandin F2α treatment at the end of an estradiol-based protocol for timed artificial insemination in lactating dairy cows during cool or hot seasons of the year. J Dairy Sci 2015;98:947–59.
- [30] Beam SW, Butler WR. Energy balance and ovarian follicle development prior to the first ovulation postpartum in dairy cows receiving three levels of dietary fat. Biol Reprod 1997;56:133–42.
- [31] Zhou H, Song X, Briggs M, Violand B, Salsgiver W, Gulve EA, Luo Y. Adiponectin represses gluconeogenesis independent of insulin in hepatocytes. Biochem Biophys Res Commun 2005;338:793–9.
- [32] Saremi B, Winand S, Friedrichs P, Kinoshita A, Rehage J, Dänicke S, Häussler S, Breves G, Mielenz M, Sauerwein H. Longitudinal profiling of the tissue-specific expression of genes related with insulin sensitivity in dairy cows during lactation focusing on different fat depots. PLoS One 2014;9:e86211.
- [33] Reist M, Erdin D, von Euw D, Tschuemperlin K, Leuenberger H, Delavaud C, Chilliard Y, Hammon HM, Kuenzi N, Blum JW. Concentrate feeding strategy in lactating dairy cows: metabolic and endocrine changes with emphasis on leptin. J Dairy Sci 2003;86:1690–706.
- [34] Huzzey JM, Veira DM, Weary DM, von Keyserlingk MAG. Prepartum behavior and dry matter intake identify dairy cows at risk for metritis. J Dairy Sci 2007;90:3220–33.

- [35] Roche JR, Burke CR, Meier S, Walker CG. Nutrition x reproduction interaction in pasture-based systems: is nutrition a factor in reproductive failure? Anim Prod Sci 2011;51:1045–66.
- [36] Eicher SD, Schutz M, Kearney F, Willard S, Bowers S, Gandy S, Graves K. Prepartum milking effects on parlour behaviour, endocrine and immune responses in Holstein heifers. J Dairy Res 2007;74:417–24.
- [37] Ouchi N, Kihara S, Funahashi T, Matsuzawa Y, Walsh K. Obesity, adiponectin and vascular inflammatory disease. Curr Opin Lipidol 2003;14:561–6.
- [38] Kadowaki T, Toshimasa Y, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. J Clin Investig 2006;116:1784–92.
- [39] Santos JE, Rutigliano HM, Sá Filho MF. Risk factors for resumption of postpartum estrous cycles and embryonic survival in lactating dairy cows. Anim Reprod Sci 2009:110:207–21.
- [40] Villa-Godoy A, Hughes TL, Emery RS, Chapin LT, Fogwell RL. Association between energy balance and luteal function in lactating dairy cows. J Dairy Sci 1988:71:1063—72
- [41] Tenhagen BA, Drillich M, Heuwieser W. Analysis of cow factors influencing conception rates after two-timed breeding protocols. Theriogenology 2001;56:831–8.

- [42] Lucy MC. Reproductive loss in high-producing dairy cattle: where will it end? J Dairy Sci 2001;84:1277—93.
- [43] Sangsritavong S, Combs DK, Sartori R, Armentano LE, Wiltbank MC. High feed intake increases liver blood flow and metabolism of progesterone and estradiol-17beta in dairy cattle. J Dairy Sci 2002;85:2831–42.
- [44] Van Saun RJ. Metabolic profiling and health risk in transition cows, pp. 212–213. In: Proceedings 37th annual American association of bovine practitioners convention, ft. Worth, Texas, september 2004; 23–25.
- [45] Janovick Guretzky NA, Carlson DB, Garrett JE, Drackley JK. Lipid metabolite profiles and milk production for Holstein and Jersey cows fed rumen-protected choline during the periparturient period. J Dairy Sci 2006;89: 188–200.
- [46] Lucy MC. Regulation of ovarian follicular growth by somatotropin and insulinlike growth factors in cattle. J Dairy Sci 2000;83:1635–47.
- [47] Beam SW, Butler WR. Effects of energy balance on follicular development and first ovulation in postpartum dairy cows. J Reprod Fertil 1999;54(Suppl): 411–24.
- [48] Hansen PJ. Effects of heat stress on mammalian reproduction. Philos Trans R Soc Lond B Biol Sci 2009;364:3341–50.